

# A REVIEW PRESENCE OF PLANT GROWTH PROMOTING MICROORGANISM IN CACTI PLANT

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**Abstract:** Soil is a dynamic living matrix that is essential for all life processes and not just for producing agricultural products. The ecology is greatly harmed by using chemical fertilizers in agriculture to increase yields and control weeds, pests, and diseases. An understanding of the cooperative behaviours of plants and rhizosphere microbial communities has been developed due to current public concerns about the symptoms of agrochemicals. Therefore, there is a need for biological agents that are known around the world. Plant Growth Promoting Rhizobacteria (PGPR) provides a superior solution to this issue. PGPR are bacteria that can help plants respond more effectively to biotic and abiotic challenges. Several natural elements, including soil type, plant cultivar, environmental change, anthropogenic activities, and more, impact rhizosphere microbiomes, which have been shown to enhance plant development and productivity. Rhizobacteria that fix nitrogen in a free-living and endophytic state can help achieve this goal. To improve both above-ground and underground biomass, *Rhizobium*, *Pseudomonas*, *Azospirillum*, and *Bacillus* have been found to affect crops. These organisms may therefore play an important role in achieving sustainable agribusiness results. As a result, it's critical to take this rhizosphere microbiome into account using increasingly sophisticated culture-free techniques. It emphasizes the need to investigate variables that can alter the rhizosphere microbiome and focus on the contributions of nitrogen-fixing microorganisms to productive horticultural outcomes at a reasonable cost and the methods that can be applied to improve them. Future research into rhizosphere science will be based on the advancement of sub-atomic and biotechnological methods to deal with the growth in our understanding of rhizosphere science and to achieve coordinated management of the soil microbial population. *Acinetobacter*, *Alcaligenes*, *Azospirillum*, *Azotobacter*, *Bacillus*, *Beijerinckia*, *Burkholderia*, *Derxia*, *Enterobacter*, *Gluconacetobacter*, *Herbaspirillum*, *Ochrobactrum*, *Pseudomonas*, *Rhodococcus*, *Serratia*,. Most Cactaceae species have modified axillary buds termed areoles, which are axillary buds with spines and lack leaves. The cacti are mostly succulent and have evolved to thrive in extremely xeric environments. Numerous varieties are currently grown extensively worldwide, largely as ornamentals and curiosities, due to their frequently unusual structures and appearances. Some cacti species are employed as hedge plants, fruit and fodder sources, and food for livestock. *Opuntia ficus-indica* (Linnaeus) Miller, commonly called "prickly pear" or "cactus pear," is the most widespread and economically significant species in the Cactaceae. Numerous locations cultivate this species.

**Keywords:** Rhizospheric soil, IAA, N<sub>2</sub> fixation, PSBs, *Bacillus*, EPS, Drought area, PGPR, siderophores, Cacti Plant, Hydroponic

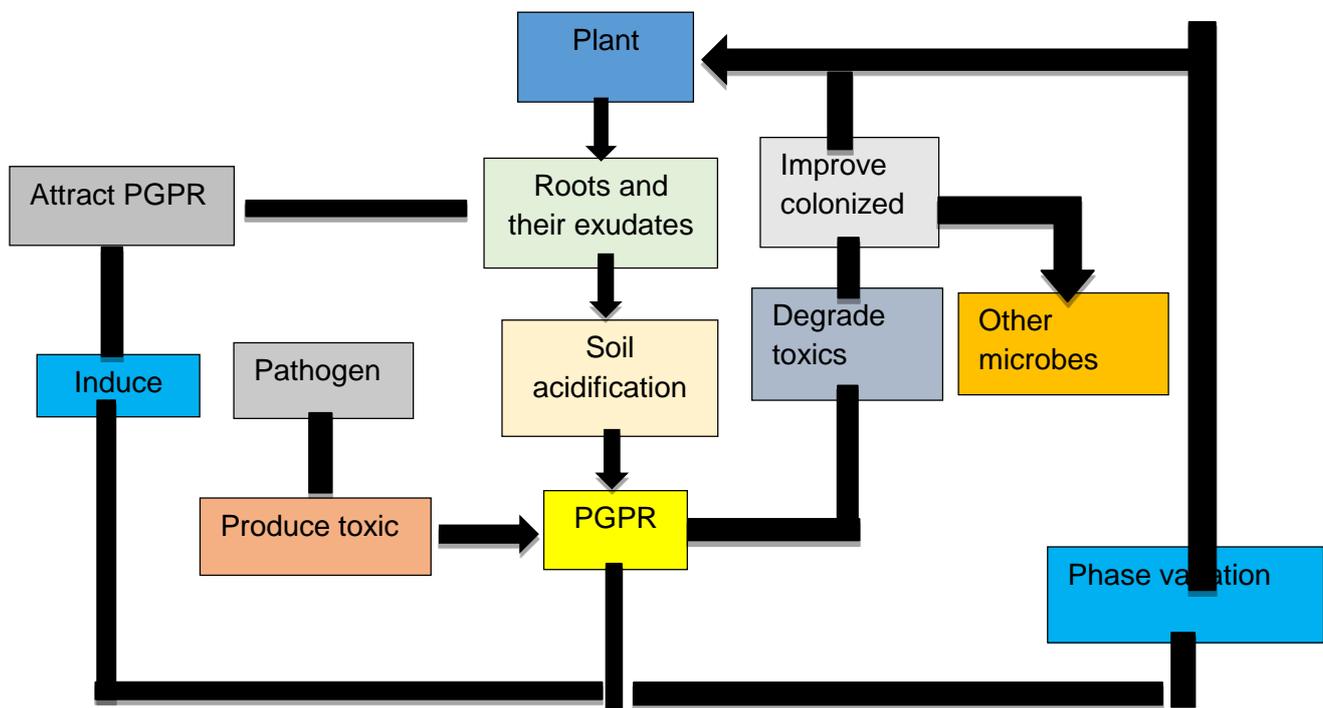
## INTRODUCTION

Cactus, plural cacti or cactuses seed plant family with nearly 2,000 species and 139 genera. Basic anatomical features of Cactaceae have been studied since the 16th century [57, 22]. More recently, other features have been observed for cultivated plants. Eight genera and 318 endemic species belonging to 42 families were listed within the Biome. Boosfeld [14] was one of the primary to emphasize the correlation of internal anatomy with external forms may also have very different external forms can also have very different internal structures. Among the modifications accompanying the evolution of cacti from a leafy ancestor that employs C<sub>3</sub> photosynthesis to stem photosynthesis. Crassulacean acid metabolism (CAM) succulent is a stem with a rise in stomatal frequency. Recent research studies have shown that microorganisms can affect the plant since they produce compounds like phytohormones and siderophores [74].

Furthermore, microbes act as phosphate solubilizers and N<sub>2</sub>-fixers and are antagonists against fungal pathogens [74, 44, 64]. These characteristics are directly or indirectly involved in plant growth promotion [40]. 40% of the carbon fixed by the plant is changed over into root exudates [52] which contains particle, free oxygen, water, enzyme and carbon-based compound, and root-derived compounds among microorganisms. The Association of plants with beneficial microbes, collectively called plant growth-promoting Rhizobacteria (PGPR), includes nitrogen-fixing bacteria, mycorrhizal fungi and biocontrol agents, which are bunch emphatically collaborating living beings. Negative connection plant with pathogenic microorganisms or growths. The nodule is the plant part where the bacteria, once transformed into a bacteroid, carry out the biological process [40]. Cacti occur within the new world from western and southern Canada. Genus *Rhipsalis* has scattered normally, without a doubt, by birds to tropical Africa and Madagascar and across to country and Southern India [62].

**Table 1: Types of Cacti:[45]**

Common name	Scientific name	Soil pH	Special Characteristics
Acanthocalycium glaucum	<i>Acanthocalycium glaucum</i>	6.1-7.8	Container, showy flowers.
Armatocereas Oligogonus	<i>Armatocereas Oligogonus</i>	6.1-7.8	Showy flowers.
Arrojadoa penicillata	<i>Arrojadoa penicillata</i>	6.1-7.8	Container
Austrocylindropuntia	<i>Austrocylindropuntia</i>	6-8	Container
Ball cactus	<i>Parodia Magnifica</i>	6.1-7.8	Container, Tolerates drought and is easy to grow.
Beaver Tail cactus	<i>Opuntia basilaris</i>	6.1-7.8	Container, Tolerates drought.
Brain cactus	<i>Stenocactus Multicostatus</i>	6.1-7.8	Container, Tolerates drought.



**Figure 1 Interaction of plant root exudates, pathogens, PGPR, and other beneficial microbes in the rhizosphere [38].**

In 1888 Beijerinck obtained the first pure bacterial culture named *Bacillus radicol. Nodule suspension*. These isolates could nodulate Pisum, and vicia was later renamed *Rhizobium leguminosarum* [27, 40] and collected the soil sample from different sides. Root portion soil collected. Add into sterile H<sub>2</sub>O and quiet down the soil after dilution and plating. Perform the gram staining and motility and another test. Isolation and screening in organism's *pseudomonas stutzeri*, *Bacillus Cereus*, *Bacillus subtilis*, *Bacillus amyloliquefaciens*, *Bacillus spp*, *pseudomonas spp*, *serratia*, *Azotobacter spp*, *microbacterium*, *S. aureus*, *E. Coli*, *Klebsiella*, *Enterobacter*.

**Table 2 Some indirect mechanisms of PGPR**

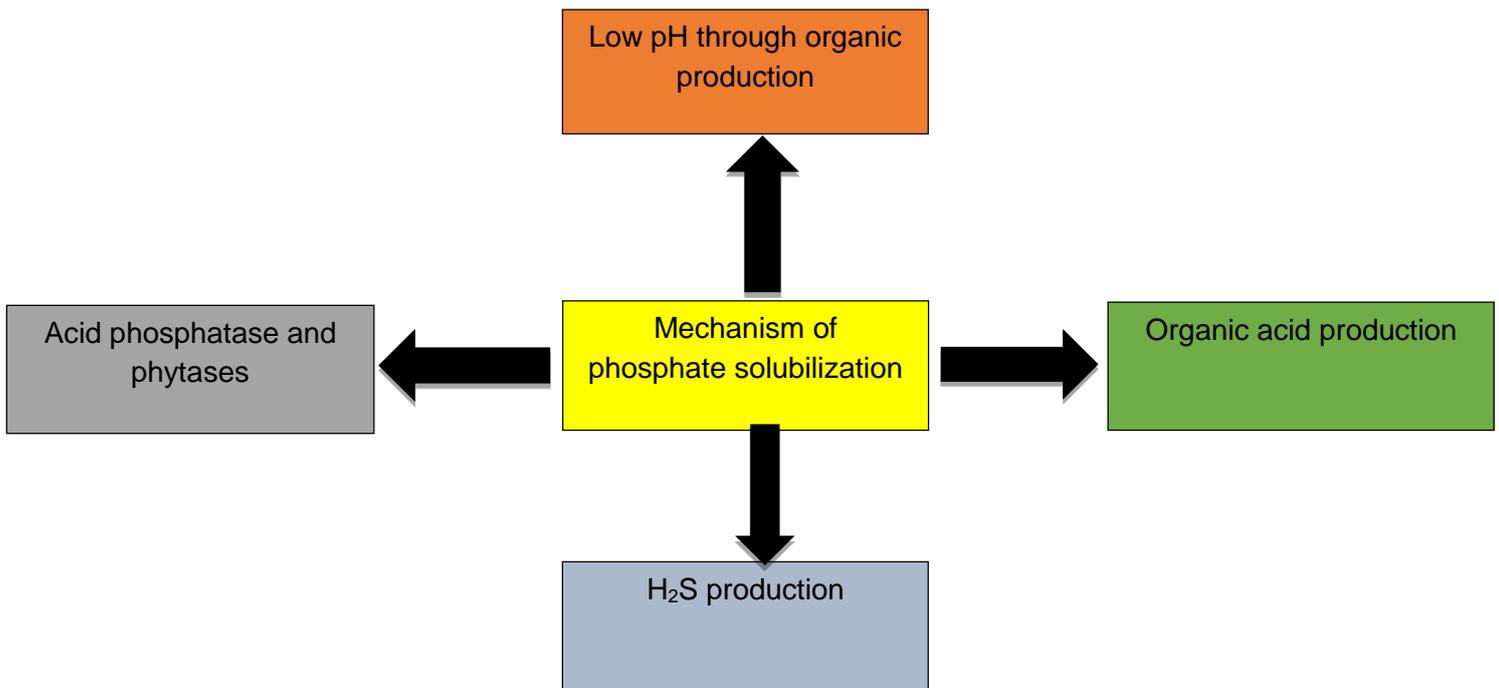
Mechanisms	Associated Micro-Organisms	Reference
Phosphate solubilization	<i>Bacillus spp.</i> <i>Enterobacterium</i> <i>microbacterium</i> <i>Pseudomonas spp.</i> <i>Rhizobium</i> <i>Serratia</i>	[77, 78]
Nitrogen fixation	<i>Azotobacter spp</i> <i>Rhizobia spp</i> <i>Azospirillum spp.</i> <i>Bacillus polymyxa</i>	[77, 78]
Production of siderophores	<i>Staphylococcus aureus</i> <i>Escherichia coli</i>	[51, 77, 78]
Indole Acetic Acid produce	<i>Klebsiella</i> <i>Enterobacter</i>	[77, 78]
Exopolysaccharides produce	<i>Rhizobacteria</i>	[67, 77, 78]

### I. INORGANIC PHOSPHATE SOLUBILIZATION

Plant development advancing rhizobacteria are the dirt microorganisms that colonized plant roots. Work with the plant development straightforwardly or by implication. Decrease the world's dependence on hazardous agricultural chemicals. Animate plant development through activating supplements in soils. Proficient at colonizing the foundation surface to survive, multiply and compete with other microbiota. Promote plant growth classification supported their functional activity as Biofertilizer, phytostimulators, Rhizoremediators, and Biopesticides. Plant development advancing rhizobacteria advance plant development straightforwardly and by implication [77].

Phosphate-solubilizing microbes utilize different mechanisms to achieve the insoluble types of phosphate into dissolvable structures. Natural acids delivered by the miniature life forms go about as great chelators of divalent cations of Ca<sup>2+</sup> going with the arrival of phosphates from insoluble phosphatic compounds. Natural acids may frame dissolvable buildings with metal particles related to insoluble 'P', delivering the phosphate [36]. A considerable lot of the PSMs bring down the pH of the medium either by H<sup>+</sup> expulsion [36] or by emission of natural acids, for example, acidic, lactic, malic, succinic, tartaric, gluconic, 2-ketogluconic, oxalic and citrus extracts. The contribution of microorganisms in the solubilization of inorganic phosphates was referred to as soon as 1903 [48]. Constitute 20 to 40% of the culturable populace of soil microorganisms, and a critical extent of these can be disengaged from rhizosphere soil [18]. Most PSB are confined from the rhizosphere of different plants and are known to be metabolically more unique than those secluded from sources other than the rhizosphere. These low degrees of P are because of the great reactivity of dissolvable P with calcium (Ca), iron (Fe) or aluminium (Al), which prompts P precipitation. Inorganic P in acidic soils is related to Al and Fe compounds. However, calcium phosphates are the dominating type of inorganic phosphates in calcareous soils. Natural P may likewise make up an enormous part of solvent P, however much half in soils with high natural matter substance [10].

Soil phosphates are delivered accessible either by plant roots or by microorganisms. Therefore, phosphate-dissolving soil microorganisms play an important role in correcting the phosphorus deficiency of crop plants [87]. Pikovskayas Agar was modified by Sundara Rao and Sinha [88, 82] to detect phosphate-solubilizing bacteria from soil.



**Figure 2 Mechanism of phosphate solubilizing[24].**

## II. ASYMBIOTIC ORGANIC PROCESS

A Symbiotic biological process fixes atmospheric nitrogen and supplies it to plants by two mechanisms. I. Symbiotic organic process. The mutual relationship between microbe and the plant. II. Non-symbiotic nitrogen fixation: Is administrated by free lifestyle diazotrophs. Biological nitrogen-fixing PGPR helps in disease management and growth-promoting activity and maintain nitrogen level in agricultural soil [77].

Rhizosphere-related N-fixing microbes have progressively been utilized in non-vegetable yield species, for example, Sugar beet, Sugar stick, Rice, Jatropha, Maize, and Wheat [76]. For instance, exploring different avenues regarding *Bacillus species* showed yield expansions in cereals [15] and maize [65]. They were taking advantage of PGPR's potential to upgrade crop efficiency utilized N<sub>2</sub>-fixing microorganisms, with the understood presumption that this movement delivered the improved harvest yields. One concentrate in Russia to test the capability of a kind of *A. radiobacter*, detached from the rhizosphere of rice, on winter wheat and spring grain seemed to give huge increments (5-30%) in yield in 2 out of 3 years. Simultaneously, it assessed that the commitment of N<sub>2</sub> obsession to add up to N osmosis was somewhere between 23 and 32% [9]. Such as *Enterobacter*, *Klebsiella*, *Burkholderia*, and *Stenotrophomonas*, have been considered in the new years in light of their relationship with significant yields and potential to improve plant development [71]. N-fixing bacterial strains *Pseudomonas putida RC06*, *Paenibacillus polymyxa RC05* and *RC14*, and *Bacillus OSU-142* have extraordinary potential and have plans; they are utilized as biofertilizers for improved yield and the nature of wheat, sugar beet, and spinach development [16]. The N-fixing *Bacillus* strains and *A. brasilense sp246* have the potential for plant development action of spring wheat and grain development in natural and low-N input farming [17].

Ashbys Agar Media are formulated [87]. It is utilized to separate *Azotobacter*, a non-harmonious nitrogen-fixing microorganism that involves mannitol as a carbon source and air nitrogen as a nitrogen source. Dipotassium phosphate provides buffering to the medium. Different fundamental particles expected to advance the development of *Azotobacter* are additionally accessible in this medium.

## III. SIDEROPHORES PRODUCTION

Siderophores could be an iron-chelating compound that helps in the assimilation of iron. Siderophores are applied in both direct and indirect enhancement of plant growth. Impact of microbially secreted siderophores on plant growth [51,77].

Siderophore is a low atomic weight compound (400-1,500 Dalton), specially chelate iron (Fe<sup>+3</sup>) and transports into the cell across the cell layer. The bacterium that initially blended the siderophores takes up the iron siderophore complex by utilizing a receptor that is well defined for the complex and is situated in the external cell layer of the bacterium. The iron is delivered and accessible to help microbial development when inside the cell. Iron is a significant micronutrient utilized by microorganisms and is fundamental for their digestion. In the dirt, it is inaccessible for direct osmosis by microorganisms because ferric iron (Fe<sup>3+</sup>), which prevails in nature, is just sparingly solvent and too low in focus to help microbial development. Soil microorganisms combine and discharge this low-sub-atomic iron restricting compound. The siderophores tie the majority of the Fe<sup>+3</sup> in the rhizosphere and successfully forestall the multiplication of infectious microorganisms by denying them accessible iron [46]. Concealment of the microbes emerges because lack of iron causes development restraint, the decline in nucleic corrosive amalgamation hindrance of sporulation, and causes changes in cell morphology [54]. Therefore, each siderophores structures a hexadentate octahedral complex with Fe<sup>+2</sup>. Catecholates are synthetically subordinates of 2, 3, dihydroxy benzoic corrosive. Each catecholates gave two oxygen particles to chelation with iron so that a hexadentate octahedral complex is shaped [20]. A horde of ecological variables can adjust the siderophore amalgamation, pH, iron level and types of iron particles, presence of minor components, and a sufficient inventory

of C, N, and P [23]. Microbial siderophores differ broadly in general construction; however, most contain hydroxamate and catechol gatherings, which are used to chelate the ferric particle [60].

Chrome azurol sulfonate (CAS) assay is one of the most used assays for detecting siderophores produced by the microbes. Cultures were point immunized on CAS agar plates and hatched at room temperature [72]. Siderophore production was studied using a succinate medium (SM) [75].

#### IV. INDOLE COMPOUNDS

Indole Carboxylic acid production Produced via way of means PGPR. Help in plant natural procedure and differentiation. Stimulates seed and tuber germination, Biosynthesis of various metabolites, and resistance to traumatic conditions. *Pseudomonas*, *Rhizobium*, *Bradyrhizobium*, *Agrobacterium*, *Enterobacter* and *Klebsiella* are IAA-generating PGPR. The colorimetric technique is employed [77].

Indole-3-acetic acid (IAA) is a critical plant chemical that governs various plant blast and improvement parts. It is the maximum not unusual place evidently to be had and physiologically vital phytohormone of the auxin class, even though numerous compounds like 4-chloroindole 3-acetic acid, phenylacetic acid, indole-3-butyric acid, and indole-3-propionic acid with comparable activity. The function of bacterial IAA is numerous in terms of the interplay among plant life and microorganisms. Previously, bacterial auxin manufacturing was believed to be especially related to pathogenesis, particularly bacterial gall formation. However, it's miles apparent that numerous phytopathogenic micro-organisms, gall-inducing in addition to plant boom selling micro-organisms, can synthesize IAA. Besides being produced employing plant life, it's also synthesized via root-related micro-organisms like *Rhizobium sp.*, *Pseudomonas sp.*, and *Azospirillum sp* [83].

The production of IAA was assayed qualitatively determined by using the Salkowski method. Tryptophan was an amino acid that plays a key role in some microorganisms' precursor production of IAA [32].

#### V. EXOPOLYSACCHARIDES

Production of exo-polysaccharides is vital in biofilm formation and root colonization. Effective colonization of plant roots by EPS-producing microbes helps carry the free phosphorous and circulating essential nutrients to the plant [67]. Functions performed by EPS-producing microbes constitute shielding from desiccation attachment to surfaces, plant invasion, and plant defence response in plant-microbe interactions [34,77].

Exopolysaccharides are starch polymers emitted by a wide assortment of plant development advancing rhizobacteria. They can remain related to the cell wall to shape a bound container layer or be delivered into cells encompassing as extracellular ooze [31]. EPS play fundamental parts in an assortment of cycles, for example, arrangement of biofilm [13], assurance of bacterial cell from desiccation, keeping up with essential cell capabilities, what's more, antibacterial movement against hunters, gelling skill, toxin corruption energy [29], bioremediation movement and plasma subbing limit.

The EPS production was checked in a modified medium by Guimarães et al. [34], a drop of 3% KOH (w / v in H<sub>2</sub>O), and then stirred repeatedly using a needle ose. Lift loopful ordinarily from the suspension surface, see whether the structure of the microorganism a tacky suspension lifted like a string with a needle ose. When the suspension turned slimy, sticky and lifted like a thread with a needle ose, meaning Gram-negative (-), on the other hand, assuming the suspension stays weakened, not rose like the needle string circle, meaning Gram-positive (+). The selected bacterium is gram-negative bacteria. To prove and validate whether these bacteria are gram-negative bacteria producing exopolysaccharides grown on the Macconkey medium is characterized by the presence of slime formed [73].

#### VI. IN VITRO ROOT COLONISATION

Surface disinfectant maize seed (alcohol, sodium hypochlorite, convective washed in sterile distilled water), Transfer to check tubes contributing, and keep two plates per pot. Experiments were founded to check the power of respective bacteria to induce maize growth and water deficit tolerance. A capacity of 80% was used.

#### VII. HYDROPONIC

PGPRs are primarily used in horticulture and agriculture, with forestry and phytoremediation applications growing in importance [52]. Several PGPR formulations are offered as commercial goods for agricultural productivity and are divided into two categories. Categories of biocontrol and biofertilizer goods. Frequently, combinations fall under the first category, comprising many bacteria, the latter being frequently one species. However, not much is done to determine whether these PGPR combinations are better for single species; elucidating synergistic effects will take a lot of investigation [46]. In a review of the bacterial species employed and examined over the last 30 years, Lucy et al. Count 49 out of 80 years, suggesting that most applications took place in field trials. (as illustrations) over the greenhouse, and as anticipated from application in soils, permit a better input-output connection. Hydroponics offers the chance to regulate microbial communities and nutrient availability compared to soil. The more direct cause of stress relief's success to the single PGP-cause-and-effect bacterium's interaction. Even the persistence of the It is possible to retain beneficial organisms at optimal densities for a longer time or with less effort adjusted. Whenever specific information on a single bacterial feature or function is available, their use in controlled settings can, for instance, release salinity tension. Advised for more efficacy.

#### CONCLUSION

The search for plant growth-promoting phosphate solubilizing bacteria is isolated for use for more than economically important crops. Phosphate solubilizing bacteria offers an environmentally sustainable approach to increasing crop production and health. Investigating that contribution to bacterial survival under adverse conditions during inoculants productions, storage, inoculation

and colonization of seeds and plants is very important. It is crucial to understand better the role of cells storage materials like PHAs, cell surface components like EPS, LPS and surface protein in enhanced resistance of bacteria to diverse stress conditions. Nitrogen-fixing microorganisms convert vaporous N from the air to inorganic blenders. Even though the job of vegetables in N obsession is obvious, the undertaking is excessively hard for them alone. The obsession interaction happens thanks to the beneficial interaction of vegetables and nitrogen-fixing microscopic organisms. It is typical for Rhizobium, colonizing vegetable roots. Be that as it may, their advantageous interaction isn't the main choice: free-living and related N obsession life forms exist. This work showed the capability of Bacillus spp. for maize development advancement. Other than this is the main report to depict desert flora-related microbes from plant development advancing capacities.

## REFERENCES

- Ahmed, E., & Holmström, S. J. (2015). Siderophore production by microorganisms isolated from a podzol soil profile. *Geomicrobiology Journal*, 32(5), 397-411.
- Ali, S. Z., Sandhya, V., Grover, M., Kishore, N., Rao, L. V., & Venkateswarlu, B. (2009). Pseudomonas sp. strain AKM-P6 enhances tolerance of sorghum seedlings to elevated temperatures. *Biology and Fertility of Soils*, 46(1), 45-55.
- Aliyat, F. Z., Maldani, M., El Guilli, M., Nassiri, L., & Ibijbijen, J. (2022). Phosphate-Solubilizing Bacteria Isolated from Phosphate Solid Sludge and Their Ability to Solubilize Three Inorganic Phosphate Forms: Calcium, Iron, and Aluminum Phosphates. *Microorganisms*, 10(5), 980.
- Aini, N., Yamika, W. S. D., & Ulum, B. (2019). Effect of nutrient concentration, PGPR and AMF on hydroponic lettuce plant growth, yield, and nutrient uptake. *Int. J. Agric. Biol*, 21, 175-183.
- Khalid, A., Arshad, M., & Zahir, Z. A. (2004). Screening plant growth-promoting rhizobacteria for improving growth and yield of wheat. *Journal of applied microbiology*, 96(3), 473-480.
- Allison AG (1998) Exopolysaccharide production in bacterial biofilm. *Biofilm J* 3(2): 1-19
- Avis, T. J., Gravel, V., Antoun, H., & Tweddell, R. J. (2008). Multifaceted beneficial effects of rhizosphere microorganisms on plant health and productivity. *Soil biology and biochemistry*, 40(7), 1733-1740.
- Babalola, O. O. (2010). Beneficial bacteria of agricultural importance. *Biotechnology Letters*, 32(11), 1559-1570.
- Bairamov LE, Vinogradova LV, Zavalin AA (2001). Nitrogen nutrition and productivity of barley as conditioned by the application of associative diazotrophs. *Asp Appl Biol* 63:135-139
- Barber SA (1984). Soil nutrient bioavailability. John Wiley, New York, USA
- Bargy's Manual of Systematic Bacteriology, Volume three (second edition) and Volume two Part-B.
- Benzerara, K., Menguy, N., López-García, P., Yoon, T. H., Kazmierczak, J., Tyliczszak, T., ... & Brown, G. E. (2006). *Nanoscale detection of organic signatures in carbonate microbialites. Proceedings of the National Academy of Sciences*, 103(25), 9440-9445.
- Bhaskar PV, Bhosle, NB (2005). Microbial extracellular polymeric substances in marine biogeochemical processes. *Curr Sci* 88(1): 45-53
- Boosfeld A. 1920 Beirrage zur vergleich enden Anatomie stammukkulenter pflanzen. *Beibungen der Botanisches central latta* 37:217-258
- Cakmakci R, Kantar F, Sahin F (2001). Effect of N<sub>2</sub>-fixing bacterial inoculations on yield of sugar beet and barley. *J Plant Nutr Soil Sci* 164:527-531
- Cakmakci UG, Erdogan, Donmez MF (2007). The Effect of plant growth-promoting rhizobacteria on barley seedling growth, nutrient uptake, some soil properties, and bacterial counts. *Turk J Agric For* 31:189-199
- Canbolat MY, Bilen S, Cakmakci R, Sahin F, Aydin A (2006). Effect of plant growth-promoting bacteria and soil compaction on barley seedling growth, nutrient uptake, soil properties and rhizosphere microflora. *Biol Fertil Soils* 42:350-357
- Chabot R, Antoun H, Cescas MP (1993). Stimulation de la croissance du maïs et de la laitue romaine par des microorganismes dissolvant le phosphore inorganique. *Can J Microbiol* 39: 941- 947
- Chanway, C. P., Hynes, R. K., & Nelson, L. M. (1989). Plant growth-promoting rhizobacteria: effects on growth and nitrogen fixation of lentil (*Lens esculenta* Moench) and pea (*Pisum sativum* L.). *Soil Biology and Biochemistry*, 21(4), 511-517.
- Chincholkar SB, Chaudhari BL, Talegaonkar SK, Kothari RM (2000). Microbial iron chelators: A sustainable tool for the Biocontrol of plant diseases. In: *Biocontrol potential and its exploitation in sustainable agriculture*, Kluwer academic press, pp. 49-69
- Choudhary, D. K., Johri, B. N., & Prakash, A. (2008). Volatiles as priming agents that initiate plant growth and defence responses. *Current Science*, 595-604.
- Conde, L. F. (1975). Anatomical comparisons of five species of *Opuntia* (Cactaceae). *Annals of the Missouri Botanical Garden*, 425-473.
- Duffy BK, Defago G (1999). Environmental factors modulating antibiotic and siderophore biosynthesis by *Pseudomonas fluorescens* biocontrol strains. *Appl Environ Microbiol* 65:2429-2438
- Etesami, H., & Adl, S. M. (2020). Plant growth-promoting rhizobacteria (PGPR) and their action mechanisms in the availability of nutrients to plants. *Phyto-Microbiome in stress regulation*, 147-203.
- Experimental Microbiology Volume 1 and 2 of Rakesh J Patel and Kiran S Trivedi.
- Figueiredo, M. D. V. B., Bonifacio, A., Rodrigues, A. C., & Araujo, F. F. D. (2016). Plant growth-promoting rhizobacteria: key mechanisms of action. In *Microbial-mediated induced systemic resistance in plants* (pp. 23-37). Springer, Singapore.

27. Frank B(1989)Ueber die Pilsymbiose der Leguminosen.BotGes 7:332-346
28. Funaki, H., & Tasaka, M. (2009). *Hormone interactions during lateral root formation*. *Plant molecular biology*, 69(4), 437-449.
29. Fusconi R, Godinho MJL (2002). Screening for exopolysaccharide producing bacteria from sub- tropical polluted groundwater. *Braz J Biol* 62(2): 363-369
30. Gibbon, A. C. 1988a.The systematics and evolution of subtribe stenocereinae. 3 Myrtillocactus. *Cactus and Succulent Journal(U. S)* 60:283-288
31. Gilck BR, Patten CL, Holguin G, and Penrose DM (1999). Biochemical and genetic mechanisms used by plant growth-promoting bacteria Imperial college press, London, pp.187-189
32. G.N. Cohen, *Microbial Biochemistry: Third edition*, (Springer, New York, 2014), p.434.
33. Gordon, S.A.; Weber, R.P. Colorimetric estimation of indoleacetic acid. *Plant Physiology*, v.26, n.1, p.192-195, 1951. <https://doi.org/10.1104/pp.26.1.192>
34. Guimarães, D.P.; Costa, F.; Rodrigues, M.J.; Maugeri, F. Optimization of dextran synthesis and acidic hydrolysis by surface response analysis. *Brazilian Journal of Chemical Engineering*, v.16, n.2, p.129-139,1999.
35. H.R.; Sandoval, A.P.S.; Oliveira, L.M.; Souza, J.T.; Soares, A.C.F. Diazotrophic bacteria associated with sisal (*Agave sisalana*Perrine ex Engelm): potential for plant growth promotion. *Plant and Soil*, v.385,p.37-48, 2014. <https://doi.org/10.1007/s11104-014-2202-x>.
36. Illmer P, Schinner F (1995). Solubilization of inorganic calcium phosphates-solubilization mechanisms. *Soil Biol Biochem* 27: 265–270
37. Jay Prakash Verma, Janardan Yadav, Kavindra Nath Tiwari, Durgesh Kumar Jaiswal. Evaluation of plant growth-promoting activities of microbial strains and their Effect on growth and yield of chickpea (*Cicer arietinum* L.) in India. *Soil Biol. Biochem.*, 70: 33-37. (2014)
38. Jha, C. K., & Saraf, M. (2015). Plant growth-promoting rhizobacteria (PGPR). *J. Agric. Res. Dev*, 5, 108-119.
39. Kavamura, V. N., Santos, S. N., da Silva, J. L., Parma, M. M., Ávila, L. A., Visconti, A., ... & de Melo, I. S. (2013). *Screening of Brazilian cacti rhizobacteria for plant growth promotion under drought*. *Microbiological Research*, 168(4), 183-191.
40. Khan, M. S. (2010). *Microbes for Legume Improvement* (A. Zaidi, M. S. Khan, & J. Musarrat, Eds.). Springer Vienna.
41. Khan, M.S., Zaidi, A., Wani, P.A. Role of phosphate-solubilizing microorganisms in sustainable agriculture- A review. *Agron. Sustain. Dév.*, 27: 29-43.(2007)
42. Khandelwal, V., Mohamed, M. N., Shukla, A. K., Mangalassery, S., & Dayal, D. (2019). Establishment and Performance of Cactus (*Opuntia ficus-indica*) Accessions at Initial Stages under Shed Net in Semi-Arid Region of Rajasthan. *Int. J. Curr. Microbiol. App. Sci*, 8(10), 1983-1988
43. .Kim, K.Y., Jordan, D. and Mc Donald, G.A. Effect of phosphate solubilizing bacteria and vesicular-arbuscular mycorrhizae on tomato growth and soil microbial activity. *Biol. Fertil. Soils*, 26: 79-87. (1998)
44. Kim, S., Lowman, S., Hou, G., Nowak, J., Flinn, B., & Mei, C. (2012). *Growth promotion and colonization of switchgrass (Panicum virgatum) cv. Alamo by bacterial endophyte Burkholderia phytofirmans strain PsJN*. *Biotechnology for Biofuels*, 5(1), 1-10.
45. Keen, B. (2011). *Cacti and Succulents: Step-by-step to Growing Success*. Crowood.Kloepper JW, Leong J, Teintze M, Schroth MN (1980). Enhanced plant growth by plant growth-promoting rhizobacteria. *Nature* 286:885–886
46. Kloepper, J.W., Ryu C.-M. and Zhang, S. (submitted). Induced systemic resistance and promotion of plant growth by *Bacillus* spp.
47. Kokalis-Burelle, N., Vavrina, C.S., Roskopf, E.N. and Shelby, R.A. 2002. Field evaluations of plant growth-promoting Rhizobacteria amended transplant mixes and soil solarization for tomato and pepper production. *Plant and Soil* 238, 257-266.
48. Kucey RMN, Jenzen HH, Leggett ME (1989). Microbially mediated increases in plant-available phosphorus. *Adv Agron* 42: 199-228
49. Kulasooriya, S. A., Seneviratne, G., & Ekanayake, E. M. H. G. S. (2017). Soil Microbial Diversity and Its Utilization in Agriculture in Sri Lanka. In *Microbial Biotechnology* (pp. 203-224). Springer, Singapore.
50. Kumari, A., Patel, A. K., Banjare, U., & Pandey, K. D. (2020). GROWTH PROMOTING CHARACTERISTICS OF ENDOPHYTIC BACTERIA ISOLATED FROM *COSTUS IGNEUS* PLANT. *Plant Archives*, 20(1), 2991-3001.
51. Loudon, B.C.; Haarmann, D.; Lynne, A.M. Use of blue agar CAS assay for siderophore detection. *Journal of Microbiology & Biology Education*, v.12, n.1, p.51-53, 2011.
52. Lucy, M., Reed, E. and Glick, B.R. 2004. Applications of free living plant growth-promoting rhizobacteria. *Antonie van Leeuwenhoek* 86, 1-25.
53. Lynch JM Whipps JM(1990) Substrate flow in the rhizosphere. *Plant Soil* 129:1-10
54. Mathiyazhagan S, Kavitha K, Nakkeerans S, Chandrasekar MK, Renukadevi P, Krishnamoorthy AS, Fernando WGD (2004). PGPR mediated management of stem blight of *Phyllanthus amarus* (Schum and Thonn) caused by *Corynespora cassiicola* (Berk and Curt) wei. *Arch Phytopathol Plant Prot* 37:183–199.
55. Mauseth, J. O. 1996.Collapsible water storage cells on *cacti* *American Journal Botany* 80:928-932)
56. *Mishra A, Salokhe VM (2008) Seedling characteristics and early growth of transplanted rice under different water regimes. ExperAgric* 44:1–19
57. Metcalf, C. R., & Chalk, L. (1950). *Anatomy of the dicotyledons*. Clarendon Press, Oxford, 2, 1014-1024.

58. Mutumba, F.A.; Zagal, E.; Gerding, M.; Castillo-Rosales, D.; Paulino, L.; Schoebitz, M. *Plant growth-promoting rhizobacteria for improved water stress tolerance in wheat genotypes. Journal of Soil Science and Plant Nutrition*, v.18, n.4, p.1080-1096, 2018.<https://doi.org/10.4067/S0718-95162018005003003>.
59. Nadeem, S. M., Ahmad, M., Zahir, Z. A., Javaid, A., & Ashraf, M. (2014). The role of mycorrhizae and plant growth-promoting rhizobacteria (PGPR) in improving crop productivity under stressful environments. *Biotechnology advances*, 32(2), 429-448.
60. Neilands JB (1995). Siderophores: structure and function of microbial iron transport compounds. *J Biol Chem* 270:26723–26726
61. Nobel P. S. 1991 Tansley Review 32, Achievable productivities of CAM Plants ;basis for high values compared with C<sub>3</sub> and C<sub>4</sub> plants. *The New Phytologist* 119:183-205
62. Nobel, P. S., & Nobel, P. o. B. a. A. D. o. t. U.-D. L. P. S. (Eds.). (2002). *Cacti: Biology and Uses*. University of California Press.
63. Nyffeler R, and U Eggli 1997. Comparative stem anatomy and *Systematic of Eriosyce Sensalato(Cactáceas)*. *Annals of Botany*80:767-786
64. Oba H, 2001 Arbuscular mycorrhizal fungi and their response to bradyrhizobial inoculation in low P. *Soil Appl soil Ecol* 13:251-258
65. Pal SS (1998) Interaction of an acid-tolerant strain of phosphate solubilizing bacteria with a few acid-tolerant crops. *Plant Soil* 198:169–177
66. Parkinson D, Gray JRG, Williams ST (1971) Methods for studying the ecology of soil microorganisms. Oxford Blackwell, Oxford
67. Paulo, E.M.; Vasconcelos, M.P.; Oliveira, I.S.; Affe, H.M.J.; Nascimento, R.; Melo, I.S.; Roque, M.R.A.; Assis, A.S. An alternative method for screening lactic acid bacteria for the production of exopolysaccharides with rapid confirmation. *Ciência e Tecnologia de Alimentos*, v.32, n.4, p.710-714, 2012. <https://doi.org/10.1590/S0101-20612012005000094>
68. Perrig, D., Boiero, M. L., Masciarelli, O. A., Penna, C., Ruiz, O. A., Cassán, F. D., & Luna, M. V. (2007). Plant-growth-promoting compounds produced by two agronomically important strains of *Azospirillum brasilense*, and implications for inoculant formulation. *Applied microbiology and biotechnology*, 75(5), 1143-1150.
69. Queiroz, B.P.V.; Aguilar-Vildoso, C.I.; Melo, I.S. Visualização in vitro da colonização de raízes por rizobactérias. *Summa Phytopathologica*, v.32, n.1, p.95-97, 2006. <https://doi.org/10.1590/S0100-5405200600010007>.
70. Ramirez LEF, Mellado JC (2005). Bacterial biofertilizers. In: Siddiqui ZA (Eds.) PGPR: Biocontrol and Bio fertilization. Springer, Dordrecht, Netherlands, pp. 143–172
71. Ramirez LEF, Mellado JC (2005). Bacterial biofertilizers. In: Siddiqui ZA (Eds.) PGPR: Biocontrol and Bio fertilization. Springer, Dordrecht, Netherlands, pp. 143–172.
72. Raval, A.A. and P.B. Desai (2015). Screening and Characterization of Several siderophore Producing Bacteria as Plant Growth-Promoters and Biocontrolling agents, *International Journal of Pharmacy and Life Sciences*, 6(10-11): 4803-4811.
73. Remel (2005). Microbiology Products: Instructions for use of MacConkey Agar. <http://www.remelinc.com>. [28 Jun 2013].
74. Ryan, R. P., Monchy, S., Cardinale, M., Taghavi, S., Crossman, L., Avison, M. B., ... & Dow, J. M. (2009). *The versatility and adaptation of bacteria from the genus Stenotrophomonas*. *Nature Reviews Microbiology*, 7(7), 514-525.
75. R. Z Sayyed, N.S Gangurde, P.R Patel, S.A Joshi and S.B Chincholkar, "Siderophore production by *Alcaligenes faecalis* and its application for growth promotion in *Arachis hypogea*", *Indian Journal of Biotechnol*, 9:302-307.
76. Sahin FC, Akmac IR, Kantar F (2004). Sugar beet and barley yields in relation to inoculation with N<sub>2</sub>-fixing and phosphate solubilizing bacteria. *Plant Soil* 265:123–129
77. Santos, A. F. J., de Moraes, J. S., Miranda, J. S., Moreira, Z. P. M., Feitoza, A. F. A., Leite, J., & FERNANDES JUNIOR, P. I. (2020). *Cacti-associated rhizobacteria from Brazilian Caatinga biome induce maize growth promotion and alleviate abiotic stress. Embrapa Semiárido-Artigo em periódico indexado (ALICE)*.
78. Santos, A.F.J.; Martins, C.Y.S.; Santos, P.O.; Corrêa, E.B.C.; Barbosa,
79. Satyaprakash, M., Nikitha, T., Reddi, E. U. B., Sadhana, B., & Vani, S. S. (2017). *Phosphorous and phosphate solubilizing bacteria and their role in plant nutrition. International Journal of Current Microbiology and Applied Sciences*, 6(4), 2133-2144.
80. Segev, E., Smith, Y., & Ben-Yehuda, S. (2012). RNA dynamics in aging bacterial spores. *Cell*, 148(1-2), 139-149.
81. Silva, S.R.; Zappi, D.; Taylor, N.; Machado, M. (Orgs.). Plano de ação nacional para a conservação das Cactáceas. Brasília:ICMbio, 2011. 112p. (Série Espécies Ameaçadas, 24). [https://www.icmbio.gov.br/portal/images/stories/doc-plano-de-acao/pan\\_cactaceas/livro\\_cactaceas\\_web.pdf](https://www.icmbio.gov.br/portal/images/stories/doc-plano-de-acao/pan_cactaceas/livro_cactaceas_web.pdf). 10 Jan. 2020.
82. Singh, Deepti, and N. Sharma. "Effect of chromium on seed germination and seedling growth of green gram (*Phaseolus Aureus* L) and chickpea (*Cicer Arietinum* L)." *Int J App Nat Sci* 6 (2017): 37-46.
83. Spaepen, S., Vanderleyden, J. Remans, R. (2007). Indole-3-acetic acid in microbial and microorganism-plant signaling. *FEMS Microbiology Reviews*. 31:425-448. DOI: 10.1111/j.1574-6976.2007.00072.x.
84. Speirs, D.C. 1982. The Cacti of western Canada (part 3) *National cactus and succulent journal* 37:53-54
85. Srinivasan, M., Holl, F. B., & Petersen, D. J. (1996). Influence of indoleacetic-acid-producing *Bacillus* isolates on the nodulation of *Phaseolus vulgaris* by *Rhizobium etli* under gnotobiotic conditions. *Canadian Journal of Microbiology*, 42(10), 1006-1014.
86. Stevenson, A., & Hallsworth, J. E. (2014). Water and temperature relations of soil A actinobacteria. *Environmental microbiology reports*, 6(6), 744-755.
87. Subba Rao N. S., 1977, *Soil Microorganisms and Plant Growth*, Oxford and IBH Publishing Co., New Delhi.

88. Sundara Rao W. V. B. and Sinha M. K., 1963, Ind. J., Agric. Sci., 33:272.
89. Sumaiya A. Shaikh, Pooja V Patel, Neelam Y.Kadam and Parimal M.Patel , Plant Growth Promoting Rhizobacteria (PGPR) An Alternative to Commercial Fertilisers.(2021).Int J Pharm Sci.12(2), b124-135.
90. Tan, X., Calderon-Villalobos L.I.A., Sharon, M., Zheng, C., Robinson, C.V., Estelle, M., Zheng, N. (2007). Mechanism of auxin perception by the TIR1 ubiquitin ligase. *Nature*. 446: 640- 645. DOI: 10.1038/nature05731.
91. Turner,B.L.,1973.chemosystematic data:their use in the study of disjunction Institute Press, Washington, D. C. Pp 27-47
92. Vavrina, C.S. 1999. The effects of (*Bacillus pumilus*) on plant growth promotion and systemic acquired resistance in muskmelon and watermelon transplants and subsequent field performance. *Proc. Int. Symp. Stand Establishment* 107-111.
93. Vejan, P., Abdullah, R., Khadiran, T., Ismail, S., & Nasrulhaq Boyce, A. (2016). Role of plant growth-promoting rhizobacteria in agricultural sustainability—a review. *Molecules*, 21(5), 573.
94. Villegas, J. and J.A. Fortin. Phosphorus solubilization and pH changes as a result of the interactions between soil bacteria
95. Whipps, J.M. 2001. Microbial interactions and Biocontrol in the rhizosphere. *Journal of Experimental Botany* 52: 487-511
96. Woitke, M., & Schitzler, W. H. (2004, November). Biotic stress relief on plants in hydroponic systems. In *International Symposium on Soilless Culture and Hydroponics* 697 (pp. 557-565).
97. Wuryanto, S., Antonius, S., & Mangunwardoyo, W. (2018, September). Production of Indole-3-acetic acid (IAA) by isolate bacteria of TPK5b2: Various of pH medium conditions. In *AIP Conference Proceedings* (Vol. 2014, No. 1, p. 020138). AIP Publishing LLC.
98. Xie, H., Pasternak, J.J. and Glick, B.R. 1996. Isolation and characterization of mutants of the plant growth-promoting rhizobacterium *Pseudomonas putida* GR12-2 that overproduce indoleacetic acid. *Current Microbiology* 32, 67-71.
99. Yoon, S. H., Ha, S. M., Kwon, S., Lim, J., Kim, Y., Seo, H., & Chun, J. (2017). Introducing EzBioCloud: a taxonomically united database of 16S rRNA gene sequences and whole-genome assemblies. *International journal of systematic and evolutionary microbiology*, 67(5), 1613.
100. Zang, F., Dashti, N., Hynes, R.K. and Smith, D.L. 1996. Plant growth-promoting rhizobacteria and soybean *Glycine max* (L.) Mar. Nodulation and nitrogen fixation at suboptimal root zone temperatures. *Annals of Botany* 77, 453-459.
101. Zhang, S., Reddy, M.S. and Kloepper, J.W. 2004. Tobacco growth enhancement and blue mold disease protection by rhizobacteria: relationship between plant growth promotion and systemic disease protection by PGPR strain 90-166. *Plant and Soil* 262: 277-288.