

A Review of Pathogenic Organisms that might contribute to Nosocomial Infection

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ABSTRACT:-

Patients receiving medical care may get nosocomial infections or infections related to healthcare. Both wealthy and poor nations around the world are affected by these illnesses. In industrialized countries, nosocomial infections account for 7% of cases, while they account for 10% in poor nations. These infections cause a prolonged stay, incapacity, and financial hardship because they happen while you're in the hospital. Infections that are frequently encountered include pneumonia brought on by a ventilator, urinary tract infections brought on by catheters, and infections at surgical sites. Bacteria, viruses, and fungal parasites are some examples of nosocomial pathogens. According to WHO estimates, these illnesses affect about 15% of all hospitalized patients. During their stay in the hospital, patients are exposed to pathogens from a variety of sources, including the environment, medical personnel, and other sick people. To prevent these illnesses, transmission should be limited. Hospital waste is a possible source of germs, with 20–25 percent of it being classified as hazardous. Nosocomial infections can be prevented by implementing infection control procedures, monitoring antibiotic use and resistance, and establishing antibiotic control policies. Effective surveillance systems can contribute both domestically and internationally. All parties must make an effort to stop and manage nosocomial infections.

Keywords: nosocomial infection, types, persist an inanimate surface

I. INTRODUCTION

Nosocomial infections (NIs) have become more common as a result of the operating room's (OT) microbial contamination [1-4,5]. It has a significant clinical impact on the patient and the compassionate surgical team [2,4,6]. With multi-drug resistant strains like methicillin-resistant Staphylococcus aureus (MRSA) [5], about 10% of all infections can have major repercussions in terms of increased patient mortality, morbidity, duration of hospital stay, and overall expenses among patients in for post-operative surgery. Antimicrobial resistance causes an increase in disease, fatalities, and medical expenses. Hospital infection control issues are getting worse due to the rise of multidrug-resistant bacteria, which are more common in underdeveloped nations and are linked to high rates of NIs and antibiotic resistance [8]. Unfiltered air, ventilation systems, antiseptic solutions, drainage of wounds, patient transportation, and collection bags, the surgical team, the volume of indoor traffic, theatre gowns, footwear, gloves, and hands, the use of inadequately sterilized equipment, the contaminated environment, and grossly contaminated surfaces have all been reported as reservoirs for OT contamination [2,4]. Depending on the number of pathogens involved, the effect of various sources on the level of microbial contamination varies. The main microorganisms connected to infection of implantable biomedical devices include Staphylococcus aureus and coagulase-negative staphylococci (CoNS), for instance [9].

By properly implementing infection control procedures, microbial contamination of the OT can be avoided. For instance, a 13-fold decrease in airborne bacteria in the OT would result in a 50% reduction in wound contamination [2]. This mostly depends on better OT cleaning, appropriate disinfection, and routine fumigation [2,4,5]. Information on the microbiological contamination of OT in Ethiopia is hard to come by. to analyze the extent of microbial contamination and determine the antibiotic resistance of the bacterial isolates from the major OT at Ayder Referral Hospital, Northern Ethiopia, this study was carried out.

The intraoperative setting increases the risk of hospital-acquired infections for Many reasons. [23-29] This shows that anesthetic practice in general, may also be connected to the emergence of hospital-acquired infections, especially in light of research showing that general anesthesia is linked to immunological suppression.[30,31] There is more pressure to create preventative measures now that this issue is becoming more widely known in the community and those pay-for-performance policies will soon be implemented. This approach might be aided by a deeper comprehension of the fundamental mechanisms governing the bacterial transmission and the rise in resistance. The anesthetic work area's medical equipment and aerosolized particles are both parts of the intraoperative environment. There is no concrete evidence connecting these parameters with the direct transmission of bacterial organisms to patients, even though this is potentially connected to the emergence of nosocomial illnesses. [23, 26,28,32]

II. NOSOCOMIAL INFECTION TYPES

The most common types of infections include pneumonia linked to ventilators, urinary tract infections linked to catheters, bloodstream infections linked to central lines, and surgical site infections. Below is a description of these:

Central line-associated bloodstream infections (CLABSI)

It has a fatality incidence rate of between 12 and 25 percent [8]. They are fatal nosocomial infections. to provide fluid and medications, catheters are put in central lines. However, extended use of these devices can result in significant bloodstream infections, which affect health and raise healthcare costs [9]. Despite a 46 percent drop in CLABSI from 2008 to 2013 in US hospitals, an estimated 30,100 CLABSI cases are still reported in ICU and acute facility wards every year.

Catheter-associated urinary tract infections(CAUTI)

The most prevalent kind of nosocomial infection worldwide is CAUTI [11]. UTIs account for more than 12% of reported infections in acute care hospitals as of 2011 [12]. The patient's endogenous native microbiota is what causes CAUTIs. In contrast to the inefficient drainage from catheters, which keeps some urine in the bladder and stabilizes bacterial residence, catheters implanted within a function as a conduit for bacterial entrance [11]? Male patients may develop orchitis, epididymitis, or prostatitis from CAUTI, whereas female patients may suffer pyelonephritis, cystitis, or meningitis [12].

Surgical site infections (SSI)

which are nosocomial infections that affect 2-5% of patients who undergo surgery, and are a common complication. These are the second most frequent kind of nosocomial infections, primarily brought on by *Staphylococcus aureus* and associated with a higher risk of death [13]. The microorganisms that cause SSI are produced by the patient's endogenous microflora. Depending on the approach and surveillance criteria utilized, the incidence could reach 20%[14].

Ventilator-associated pneumonia (VAP)

Nosocomial pneumonia, or VAP, affects 9 to 27% of patients using a mechanically assisted ventilator. After tracheal intubation, often happens within 48 hours [15]. Ventilation is linked to 86 percent of nosocomial pneumonia [16]. VAP symptoms include bronchial noises, leucopenia, and fever.

III. NOSOCOMIAL PATHOGENS

Nosocomial infections are caused by bacteria, viruses, and fungus parasites. These bacteria differ based on various patient demographics, healthcare facilities, and even variations in the environment where care is provided.

• Bacteria

The most frequent pathogens that cause nosocomial infections are bacteria. Some are a part of the patient's normal flora and only cause infection when the patient's immune system becomes vulnerable to pathogens. The group of dangerous bacteria called *Acinetobacter* is what causes infections in intensive care units. It causes 80% of reported illnesses and is ingested through soil and water [18]. A commensal bacteria called *Bacteroides fragilis* is prevalent in the colon and gastrointestinal tract. When mixed with other bacteria, it can result in illnesses [19]. Due mostly to the replacement of helpful bacteria with pathogenic ones, *Clostridium difficile* can induce inflammation of the colon, which can result in antibiotic-associated diarrhea and colitis. *C.difficile* is spread from an infected patient to others by the ill-sanitized hands of medical personnel [19]. When *Enterobacteriaceae*, which are commonly found in the gut and are carbapenem-resistant, spread to other bodily areas, they can cause infections. *Escherichia coli* and *Klebsiella* species belong to the family *Enterobacteriaceae*. The defense against them becomes more challenging due to their strong resistance to carbapenem [20]. MRSA is a kind of methicillin-resistant *S. aureus* that spreads through direct contact, open wounds, and dirty hands. Sepsis, pneumonia, and SSI are brought on by it spreading from organs or the bloodstream. It has a high level of resistance to beta-lactam antibiotics [20].

• Viruses

In addition to bacteria, viruses play a significant role in nosocomial illness. According to routine monitoring, viruses are responsible for 5% of all nosocomial infections [21]. They are spread through hand-to-mouth contact, respiratory contact, and fecal-oral contact [22]. The virus-based chronic illness known as hepatitis is.

Hepatitis viruses can be spread during the administration of healthcare to both patients and staff. Unsafe injecting techniques frequently lead to the transmission of hepatitis B and C [20]. Other viruses include rotavirus, herpes simplex, HIV, influenza, and more[22].

• Parasite fungi

In people with impaired immune systems, fungus parasites operate as opportunistic pathogens that cause nosocomial infections. Infections can be brought on by *Aspergillus* spp. environmental pollution. Additionally, infections during hospital stays are brought on by *Cryptococcus neoformans* and *Candida albicans* [22]. While *Aspergillus* infections are brought on by inhaling fungus spores from contaminated air during hospital construction or renovation, *Candida* infections are brought on by the patient's own internal flora [23].

IV.NOSOCOMIAL INFECTION PREVENTION

Nosocomial infections must be prevented from the outset to control their spread because they are a substantial cause of sickness and death[22].

• Transmission from the outside

The best habitat for the pathogenic organism to thrive is one that is unhygienic. Food, water, and the air can all become polluted and spread to the people receiving medical care. Policies must be in place to guarantee that cleaning solutions are used on the walls, floors, windows, beds, bathtubs, toilets, and other medical equipment. Airborne bacterial pollution can be eliminated with proper ventilation and fresh, filtered air. It is necessary to maintain and record routine inspections of the filters and ventilation systems in general wards, surgical rooms, and ICUs. Healthcare facilities not meeting the required standards are to blame for infections that are ascribed to water. For water analysis, microbiological monitoring techniques should be applied. Separate baths must be supplied to infected patients. Food-borne infections may result from improper food management. The environment needs to be cleaned, and the food needs to fulfill minimum standards[22].

• Transmission from staff Healthcare

workers may transmit infections. Healthcare practitioners have a responsibility to participate in infection control. Staff members need to practice good personal hygiene, thus they should do so. After having contact with infected patients, it is necessary to properly decontaminate your hands using hand disinfectants. Sterilized tools and safe injection techniques should be applied. The delivery of healthcare requires the use of masks, gloves, head coverings, and/or a proper uniform [22].

V. NOSOCOMIAL PATHOGENS PERSIST ON INANIMATE OBJECTS.

The majority of papers using experimental data explored persistence on dry surfaces utilizing lab-based artificial contamination of a predetermined kind of surface. The majority of research prepared the bacteria in broth, water, or saline. Typically, viruses were created in a cell culture medium [43]. The climatic conditions are stable in terms of temperature and air humidity, which is the key benefit. Additionally, only controlled conditions—which are much simpler to guarantee in a lab—can be used to establish the impact of temperature or relative humidity [44].

Surfaces in hospitals that people touch frequently include nosocomial diseases and could spread those viruses to other patients [45–46]. A pathogen may be transferred to the hand from a contaminated surface to another hand in varying degrees. *Escherichia coli*, *Salmonella* spp., *Staphylococcus aureus* (all 100%) [47], *Candida albicans* (90%) [48], rhinovirus (61%) [49], HAV (22–33%) [50], and rotavirus (16%) [51,52] were the pathogens most successfully transmitted to hands. Viral transmission from contaminated hands can affect 14 additional patients or 5 other surfaces [53,54]. As evidenced by HAV [50,53], contaminated hands can potentially be the cause of recontaminating the surface. The compliance rate for hand hygiene among healthcare personnel is believed to be around 50% [55].

Persistence of bacteria

Staphylococcus aureus (including MRSA), *Streptococcus pyogenes*, and other gram-positive bacteria can persist for months on dry surfaces. Between multiresistant and susceptible isolates of *Staphylococcus aureus* and *Enterococcus* spp., there was generally no discernible difference in survival [56]. Such a distinction was only suggested in one investigation, but the susceptible strains showed a very short survival as such [57]. Numerous gram-negative bacteria, including *Shigella* spp., *Acinetobacter* spp., *Escherichia coli*, *Klebsiella* spp., *Pseudomonas aeruginosa*, *Serratia marcescens*, and others, can last for weeks or even months on inanimate objects. The majority of isolates from individuals with nosocomial illnesses contain these species [58]. However, several other diseases only last for a few days, like *Bordetella pertussis*, *Haemophilus influenzae*, *Proteus Vulgaris*, or *Vibrio cholerae*. Mycobacteria, such as *Mycobacterium TB* and spore-forming organisms like *Clostridium difficile*, can also endure on surfaces for long periods. Overall, it has been noted that gram-negative bacteria survive longer than gram-positive bacteria [59,60]. Most bacterial species, including *Chlamydia trachomatis* [61], *Listeria monocytogenes* [62], *Salmonella typhimurium* [62], *Pseudomonas aeruginosa* [63], *Escherichia coli* [64], or other important pathogens [65,66], exhibited better persistence in humid environments. At low humidity, only *Staphylococcus aureus* was observed to survive longer [63]. The majority of bacteria, including *Listeria monocytogenes* [62], *Salmonella typhimurium* [62], MRSA [67], corynebacteria [68], *Escherichia coli* [64,69], *Helicobacter pylori* [70], and *Neisseria gonorrhoeae* [24], all exhibited better persistence at low temperatures, such as 4°C or 6°C. There is no consistent outcome, regardless of the test content. While some researchers claim that the type of material has no bearing on persistence [72,73], others have noted that plastic has a longer persistence than steel [74,75], and yet others have observed a survival advantage with steel [76]. Other factors were rarely looked into, hence the results are uneven. Higher inocula [75], the presence of protein [60], serum [60,71], sputum [77], and the absence of dust [57] have all been associated with long persistence.

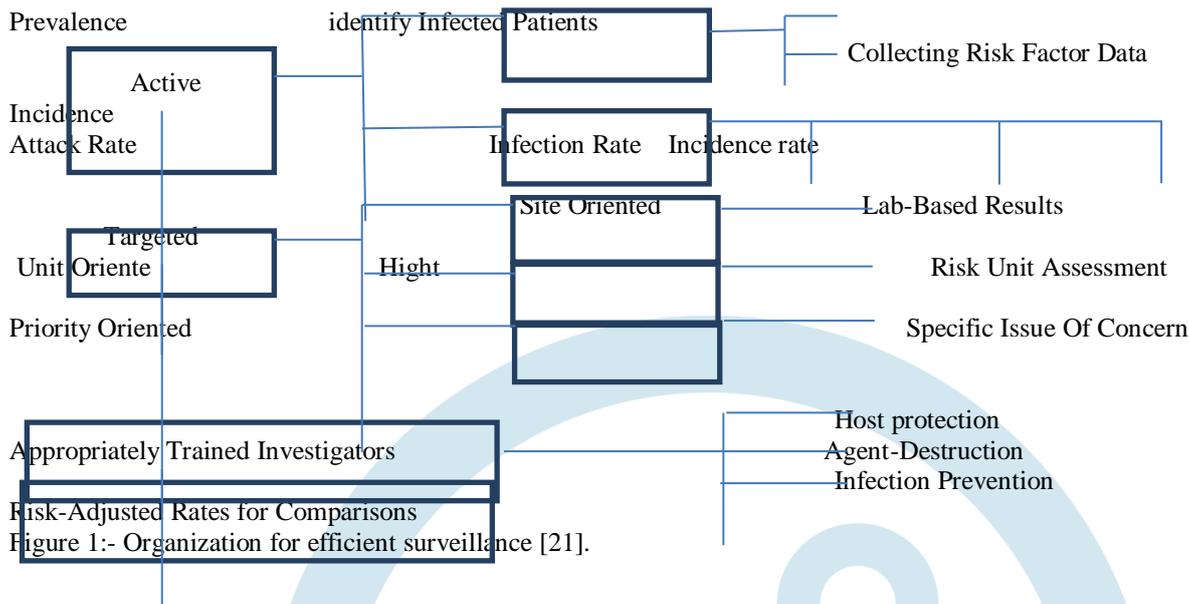
Persistence of fungi *Candida*

The most significant nosocomial fungal infection, *Candida albicans*, can live on surfaces for up to 4 months (Table 2). Other yeasts' persistence was said to be comparable to (*Torulopsis glabrata* 5 months) or shorter (*Candida parapsilosis* 14 days). Longer persistence has been attributed to the presence of serum or albumin, a low temperature, and high humidity [78].

The majority of respiratory tract viruses, including corona, coxsackie, influenza, SARS, and rhinovirus, can survive on surfaces for a few days. Astrovirus, HAV, poliovirus, and rotavirus are examples of gastrointestinal system viruses that can persist for up to two months. Blood-borne pathogens like HBV and HIV can survive for longer than a week. CMV and HSV type 1 and 2 and other herpes viruses have been shown to last from a few hours to seven days. There are conflicting descriptions of how humidity affects persistence. High humidity was linked to extended persistence for entero- [79] and rhinovirus [33]. At low humidity, HSV [34] and HAV [35] can survive longer. Conflicting results were observed for adeno- [32,34], rota- [36,37], and poliovirus [34,35]. Most viruses, including Astro- [38], adeno- [34], poliovirus [34], HSV [34], and HAV [35], are more persistent when the temperature is low. Results on the influence of material type are also said to have been inconsistent. Some authors claimed that the type of material had no impact on the persistence of norovirus [42], rotavirus [41], RSV [39], poliovirus [41], adenovirus [39-41], and echovirus [39-41].

VI. SURVEILLANCE OF NOSOCOMIAL INFECTION

Although the purpose of infection prevention and control programs is to eradicate nosocomial infections epidemiological surveillance for evidence of performance improvement is still required to reach the aim. Data collection from many sources of information by professional data collectors is one of the most effective monitoring strategies. Information should include administrative data, demographic risk factors, patients' medical histories, diagnostic testing, and data validation. After the data has been extracted, the information should be analyzed. This analysis should contain a description of the determinants, a distribution of infections, and a comparison of incidence rates. Infection control committees, management, and laboratories should communicate feedback and reports following analysis while maintaining the privacy of individuals. For the deployment of interventions to be successful and to continue, surveillance systems must be evaluated for credibility. Finally, it should become mandatory to collect data at regular intervals to maintain the effectiveness of surveillance systems [22]. The figure provides an effective way for an acceptable surveillance approach.



VII. CONCLUSION

It is now more challenging for infection control committees and healthcare administrations to achieve the goal of eliminating intervals due to rising nosocomial infection rates and antimicrobial resistance. However, it is possible to easily reduce the resistance of emerging pathogens against antibiotics by implementing sound and healthy methods for care delivery that were designed by infection control committees, controlling the transmission of these infections using appropriate methods for antibiotic use. Healthcare institutions can develop infection control plans with the aid of an effective surveillance approach supervised by WHO. Nosocomial infections can be decreased by properly training hospital staff in biosafety, waste management, and healthcare records, as well as by raising public awareness of these endemic infections.

References

- [1]. WHO. The burden of healthcare-associated infection worldwide.2016 [Online] Available from: http://www.who.int/gpsc/country_work/burden_hcai/en/ [Accessed on 10th August 2016]
- [2]. CDC. Types of healthcare-associated infections. Healthcare-associated infections (HAIs). 2016 [Online] Available from:<https://www.cdc.gov/HAI/infectionTypes.html> [Accessed on 10th August 2016]
- [3]. Raja Danasekaran GM, Annadurai K. Prevention of healthcare-associated infections: protecting patients, saving lives. *Int J Community Med Public Health* 2014; 1(1): 67-8.
- [4]. Vincent JL, Marshall J, Silva E, Anzueto A, Martin CD, Moreno R, et al. International study of the prevalence and outcomes of infection in intensive care units. *JAMA* 2009; 302(21): 2323-9.
- [5]. Allegranzi B. Report on the burden of endemic healthcare-associated infection worldwide. Geneva: WHO; 2011.
- [6]. Gupta A, Singh DK, Krutarth B, Maria N, Srinivas R. Prevalence of health care associated infections in a tertiary care hospital in Dakshina Kannada, Karnataka: a hospital-based cross-sectional study. *Int J Med Res Health Sci* 2015; 4(2): 317-21.
- [7]. Khan H, Ahmad A, Mehboob R. Nosocomial infections and their control strategies. *Asian Pac J Trop Biomed* 2015; 5(7): 509-14.
- [8]. Vital signs: Central line-associated bloodstream infections –the United States, 2001, 2008, and 2009. *Morb Mortal Wkly Rep* 2011;60(08): 243-8.
- [9]. WHO. Preventing bloodstream infections from central line venous catheters. Geneva: WHO; 2016. [Online] Available from: <http://www.who.int/patientsafety/implementation/bsi/en/> [Accessed on 10th August 2016]
- [10]. CDC. Bloodstream infection event (central line-associated blood-stream infection and non-central line-associated bloodstream infection). Atlanta, Georgia: CDC; 2015. [Online] Available from:http://www.cdc.gov/nhsn/pdfs/pscmanual/4psc_clabscurrent.pdf[Accessed on 10th August 2016]
- [11]. Warren JW. Catheter-associated urinary tract infections. *Int J Antimicrob Agents* 2001; 17(4): 299-303.
- [12]. CDC. Urinary tract infection (catheter-associated urinary tractinfection [CAUTI] and non-catheter associated urinary tractinfection [UTI]) and other urinary system infection [USI]) events.Atlanta, Georgia: CDC; 2016. [Online] Available from: <http://www.cdc.gov/nhsn/pdfs/pscmanual/7pscclauticurrent.pdf>[Accessed on 10th August, 2016]
- [13]. Anderson DJ. Surgical site infections. *Infect Dis Clin North Am*2011; 25(1): 135-53.
- [14]. Owens CD. Surgical site infections: epidemiology, microbiology, and prevention. *J Hosp Infect* 2008; 70(Suppl2): 3-10.
- [15]. Hunter JD. Ventilator-associated pneumonia. *BMJ* 2012; 344: 40-4.
- [16]. Steven M, Koenig JDT. Ventilator-associated pneumonia: diagnosis, treatment, and prevention. *Clin Microbiol Rev* 2006; 19(4):637-57.
- [17]. Hjalmarson DEC. Ventilator-associated tracheobronchitis and pneumonia: thinking outside the box. *Clin Infect Dis* 2010;51(Suppl 1): S59-66.

- [18]. Suresh G, Joshi GML. *Acinetobacter baumannii*: an emerging pathogenic threat to public health. *World J Clin Infect Dis* 2013;3(3): 25-36.
- [19]. Jayanthi A. Most common healthcare-associated infections: 25 bacteria, viruses causing HAIs, Becker's hospital review. 2014.
- [20]. CDC. Diseases and organisms in healthcare settings. Healthcare-associated infections (HAIs). Atlanta, Georgia: CDC; 2016. [Online] Available from: <https://www.cdc.gov/hai/organisms/organisms.html> [Accessed on 10th August 2016]
- [21]. Aitken CJD. Nosocomial spread of viral disease. *Clin Microbiol Rev* 2001; 14(3): 528-46.
- [22]. Duce JF, Nicolle L. Prevention of hospital-acquired infections. Geneva: WHO; 2002.
- [23]. Caffau S, Nadali L: On the bacterial contamination of anesthetic equipment. *Friuli Med* 1965; 20:515-31
- [24]. Dellinger E, Gordon S: Surgical-associated Infection in Today's Operating Room. Special Report, Anesthesiology, General Surgery, and OB/GYN News. New York, McMahan Publishing Group, 2006, pp 1-10
- [25]. Friss H, Helms P: Bacterial contamination of anesthetic equipment during use. *Ugeskr Laeger* 1963; 125:619-23
- [26]. Lessard MR, Trepanier CA, Gourdeau M, Denault PH: A microbiological study of the contamination of the syringes used in anesthesia practice. *Can J Anaesth* 1988; 35:567-9
- [27]. Leung M, Chan AH: Control and management of hospital indoor air quality. *Med Sci Monit* 2006; 12:SR17-23
- [28]. Madar R, Novakova E, Baska T: The role of non-critical health-care tools in the transmission of nosocomial infections. *Bratisl Lek Listy* 2005; 106:348-50
- [29]. Maslyk PA, Nafziger DA, Burns SM, Bowers PR: Microbial growth on the anesthesia machine. *AANA J* 2002; 70:53-6
- [30]. Hajjar J, Girard R: Surveillance of nosocomial infections related to anesthesia: A multicenter study. *Ann Fr Anesth Reanim* 2000; 19:47-53
- [31]. Rassias AJ, Marrin CA, Arruda J, Whalen PK, Beach M, Yeager MP: Insulin infusion improves neutrophil function in diabetic cardiac surgery patients. *Anesth Analg* 1999; 88:1011-6
- [32]. du Moulin GC, Saubermann AJ: The anesthesia machine and circle system are not likely to be sources of bacterial contamination. *ANESTHESIOLOGY* 1977; 47:353-8
- [33]. Sattar SA, Karim YG, Springthorpe VS, Johnson-Lussenburg CM: Survival of human rhinovirus type 14 dried onto nonporous inanimate surfaces: effect of relative humidity and suspending medium. *Canadian Journal of Microbiology* 1987, 33:802-806.
- [34]. Mahl MC, Sadler C: Virus survival on inanimate surfaces. *Canadian Journal of Microbiology* 1975, 21:819-823.
- [35]. Mbithi JN, Springthorpe VS, Sattar SA: Effect of relative humidity and air temperature on survival of hepatitis A virus on environmental surfaces. *Applied and Environmental Microbiology* 1991, 57:1394-1399.
- [36]. Ansari SA, Springthorpe VS, Sattar SA: Survival and vehicular spread of human rotaviruses: possible relation to The seasonality of outbreaks. *Reviews of Infectious Diseases* 1991, 13:448-461.
- [37]. Sattar S, Lloyd-Evans N, Springthorpe VS: Institutional outbreaks of rotavirus diarrhea: potential role of fomites and environmental surfaces as vehicles for virus transmission. *Journal of Hygiene, Cambridge* 1986, 96:277-289.
- [38]. Abad FX, Villena C, Guix S, Caballero S, Pintó RM, Bosch A: Potential role of fomites in the vesicular transmission of human astroviruses. *Applied and Environmental Microbiology* 2001, 67:3904-3907.
- [39]. Wladowetz VW, Dmitrijewa RA, Safjulin AA: Die Persistenz von Viren auf Oberflächen und die Anwendung der UV- Bestrahlung zur Virusdesinfektion. *Zeitschrift für die gesamte Hygiene* 1974, 7:173-176.
- [40]. Gordon YJ, Gordon RY, Romanowski E, Araullo-Cruz TP: Prolonged recovery of desiccated adenoviral serotypes 5, 8, and 19 from plastic and metal surfaces in vitro. *Ophthalmology* 1993, 100:1835-1839
- [41]. Abad FX, Pinto RM, Bosch A: Survival of enteric viruses on environmental fomites. *Applied and Environmental Microbiology* 1994, 60:3704-3710.
- [42]. DE'Souza DH, Williams K, Jean J, Sair A, Jaykus L: Persistence of Norwalk virus on environmental surfaces and its transfer to food Washington, D.C... *American Society for Microbiology*; 2003.
- [43]. Rzezutka A, Cook N: Survival of human enteric viruses in the environment and food. *FEMS Microbiology Reviews* 2004, 28:441-453.
- [44.] Bures S, Fishbain JT, Uyehara CF, Parker JM, Berg BW: Computer keyboards and faucet handles as reservoirs of nosocomial pathogens in the intensive care unit. *American Journal of Infection Control* 2000, 28:465-471.
- [45]. Catalano M, Quelle LS, Jeric PE, Di Martino A, Maimone SM: Survival of *Acinetobacter baumannii* on bed rails during an outbreak and sporadic cases. *Journal of Hospital Infection* 1999, 42:27-35.
- [46]. Boyce JM, Potter-Bynoe G, Chenevert C, King T: Environmental contamination due to methicillin-resistant *Staphylococcus aureus*: possible infection control implications. *Infection Control and Hospital Epidemiology* 1997, 18:622-627.
- [47]. Scott E, Bloomfield SF: The survival and transfer of microbial contamination via cloths, hands, and utensils. *Journal of Applied Bacteriology* 1990, 68:271-278.
- [48]. Rangel-Frausto MS, Houston AK, Bale MJ, Fu C, Wenzel RP: An experimental model for the study of *Candida* survival and transmission in human volunteers. *European Journal of Clinical Microbiology and Infectious Diseases* 1994, 13:590-595.
- [49]. Gwaltney JM, Hendley JO: Transmission of experimental rhinovirus infection by contaminated surfaces. *American Journal of Epidemiology* 1982, 116:828-833.
- [50]. Mbithi JN, Springthorpe VS, Boulet JR, Sattar SA: Survival of hepatitis A virus on human hands and its transfer on contact with animate and inanimate surfaces. *Journal of Clinical Microbiology* 1992, 30:757-763.
- [51]. Ward RL, Bernstein DI, Knowlton DR, Sherwood JR, Young EC, Cusack TM, Rubino JR: Prevention of surface-to-human transmission of rotaviruses by treatment with disinfectant spray. *Journal of Clinical Microbiology* 1991, 29:1991-1996.

- [52]. Ansari SA, Sattar SA, Springthorpe VS, Wells GA, Tostawaryk W: Rotavirus survival on human hands and transfer of infectious virus to inanimate and nonporous inanimate surfaces. *Journal of Clinical Microbiology* 1988, 26:1513-1518.
- [53]. Barker J, Vipond IB, Bloomfield SF: Effects of cleaning and disinfection in reducing the spread of norovirus contamination via environmental surfaces. *Journal of Hospital Infection* 2004, 58:42-44.
- [54]. von Rheinbaben F, Schunemann S, Gross T, Wolff MH: Transmission of viruses via contact in a household setting: experiments using bacteriophage strain phiXI174 as a model virus. *Journal of Hospital Infection* 2000, 46:61-66.
- [55]. Kampf G, Kramer A: Epidemiologic background of hand hygiene and evaluation of the most important agents for scrubs and rubs. *Clinical Microbiology Reviews* 2004, 17:863-893.
- [56]. Neely AN, Maley MP: Survival of enterococci and staphylococci on hospital fabric and plastic. *Journal of Clinical Microbiology* 2000, 38:724-726.
- [57]. Wagenvoort JHT, Penders RJR: Long-term in-vitro survival of an epidemic MRSA phage-group III-29 strain. *Journal of Hospital Infection* 1997, 35:322-325.
- [58]. Rüden H, Gastmeier P, Daschner FD, Schumacher M: Nosocomial and community-acquired infections in Germany. Summary of the results of the first national prevalence study (NIDEP). *Infection* 1997, 25:199-202.
- [59]. Dickgiesser N: Untersuchungen über das Verhalten grampositiver und gramnegativer Bakterien in trockenem und feuchtem Milieu. *Zentralblatt für Bakteriologie und Hygiene, I Abt Orig B* 1978, 167:48-62.
- [60]. Hirai Y: Survival of bacteria under dry conditions from a viewpoint of nosocomial infection. *Journal of Hospital Infection* 1991, 19:191-200.
- [61]. Novak KD, Kowalski RP, Karenchak LM, Gordon YJ: Chlamydia trachomatis can be transmitted by a nonporous plastic surface in vitro. *Cornea* 1995, 14:523-526.
- [62]. Helke DM, Wong ACL: Survival and growth characteristics of *Listeria monocytogenes* and *Salmonella typhimurium* on stainless steel and Buna-N rubber. *Journal of Food Protection* 1994, 57:963-968.
- [63]. Gundermann KO: Untersuchungen zur Lebensdauer von Bakterienstämmen im Staub unter dem Einfluß unterschiedlicher Luftfeuchtigkeit. *Zentralblatt für Bakteriologie und Hygiene, I Abt Orig B* 1972, 156:422-429.
- [64]. Williams AP, Avery LM, Killham K, Jones DL: Persistence of *Escherichia coli* O157 on farm surfaces under different environmental conditions. *Journal of Applied Microbiology* 2005, 98:1075-1083.
- [65]. Jawad A, Snelling AM, Heritage J, Hawkey PM: Influence of relative humidity and suspending menstrua on survival of *Acinetobacter* spp. on dry surfaces. *Journal of Clinical Microbiology* 1996, 34:2881-2887.
- [66]. Jawad A, Snelling AM, Heritage J, Hawkey PM: Exceptional desiccation tolerance of *Acinetobacter* radioresistance. *Journal of Hospital Infection* 1998, 39:235-240.
- [67]. Noyce JO, Michels H, Keevil CW: Potential use of copper surfaces to reduce survival of epidemic methicillin-resistant *Staphylococcus aureus* in the healthcare environment. *Journal of Hospital Infection* 2006, 63:289-297.
- [68]. Augustine JL, Renshaw HW: Survival of *Corynebacterium pseudotuberculosis* in axenic purulent exudate on common barnyard fomites. *American Journal of Veterinary Research* 1986, 47:713-715
- [69]. Wilks SA, Michels H, Keevil CW: The survival of *Escherichia coli* O157 on a range of metal surfaces. *International Journal of Food Microbiology* 2005, 105:445-454.
- [70]. Boehmler G, Gerwert J, Scupin E, Sinell HJ: Zur Epidemiologie der Helicobacteriose des Menschen; Untersuchungen zur Überlebensfähigkeit des Erregers in Lebensmitteln. *Deutsche Tierärztliche Wochenschrift* 1996, 103:438-443.
- [71]. Elmos T: Survival of *Neisseria gonorrhoeae* on surfaces. *Acta Dermato-Venereologica* 1977, 57:177-180.
- [72]. Wendt C, Dietze B, Dietz E, Rüden H: Survival of *Acinetobacter baumannii* on dry surfaces. *Journal of Clinical Microbiology* 1997, 35:1394-1397.
- [73]. Bale MJ, Bennett PM, Benninger JE, Hinton M: The survival of bacteria exposed to desiccation on surfaces associated with farm buildings. *Journal of Applied Bacteriology* 1993, 75:519-528.
- [74]. Pérez JL, Gómez E, Saucá G: Survival of gonococci from urethral discharge on fomites. *European Journal of Clinical Microbiology and Infectious Diseases* 1990, 1:54-55.
- [75]. Neely AN: A survey of gram-negative bacteria survival on hospital fabrics and plastics. *Journal of Burn Care and Rehabilitation* 2000, 21:523-527.
- [76]. Webster C, Towner KJ, Humphreys H: Survival of *Acinetobacter* on three clinically related inanimate surfaces. *Infection Control and Hospital Epidemiology* 2000, 21:246.
- [77]. Smith CR: Survival of tubercle bacilli: the viability of dried tubercle bacilli in unfiltered room light, in the dark, and in the refrigerator. *American Review of Tuberculosis* 1942, 5:334-345.
- [78]. Blaschke-Hellmessen R, Kreuz M, Sprung M: Umweltresistenz und natürliche Keimreservoirie medizinisch bedeutsamer Sprosspilze. *Zeitschrift für die gesamte Hygiene* 1985, 31:712-715
- [79]. Hara J, Okomator S, Minekawa Y, Yamazaki K, Kase T: Survival and disinfection of adenovirus type 19 and enterovirus 70 in ophthalmic practice. *Japanese Journal of Ophthalmology* 1990, 34:421-427